

Sustainability of Land Application of Class B Biosolids

Ian L. Pepper* and Huruy Zerzghi The University of Arizona

John P. Brooks USDA-ARS

Charles P. Gerba The University of Arizona

Land application of Class B biosolids is routinely undertaken in the United States. However, due to public concern over potential hazards, the long-term sustainability of land application has been questioned. Thus, the objective of this review article was to evaluate the sustainability of land application of Class B biosolids. To do this we evaluated (i) the fate and transport of potential biological and chemical hazards within biosolids, and (ii) the influence of long-term land application on the microbial and chemical properties of the soil. Direct risks to human health posed by pathogens in biosolids have been shown to be low. Risks from indirect exposure such as aerosolized pathogens or microbially contaminated ground water are also low. A long-term land application study showed enhanced microbial activity and no adverse toxicity effects on the soil microbial community. Long-term land application also increased soil macronutrients including C, N, and, in particular, P. In fact, care should be taken to avoid contamination of surface waters with high phosphate soils. Available soil metal concentrations remained low over the 20-yr land application period due to the low metal content of the biosolids and a high soil pH. Soil salinity increases were not detected due to the low salt content of biosolids and irrigation rates in excess of consumptive use rates for cotton. Our conclusion, based on these studies, is that long-term land application of Class B biosolids is sustainable.

THE world's 6.8 billion people all produce sewage. In the developed world most of this is treated in wastewater treatment plants, resulting in large volumes of biosolids. In the United States, the term *biosolid* implies treatment to produce Class A or Class B biosolids that meet the land application standards in the Part 503 USEPA regulations (USEPA, 1994). Currently about 60% of all biosolids in the United States are land-applied, with most of this land receiving Class B biosolids (NRC, 2002). Class B biosolids are normally produced as a "cake" (\approx 20% solids) or as liquid biosolids (\approx 5–8% solids). In either case, Class B biosolids usually result from mesophilic anaerobic digestion and, by definition, are likely to contain human pathogenic bacteria, viruses, and protozoan parasites (Pepper et al., 2006).

Although the types of land and crops to which Class B biosolids are applied is controlled, public concern in some areas of the United States, such as parts of California, has resulted in the land application of Class B biosolids being banned. This has raised the issue of the sustainability of land application in general and application of Class B material in particular. In some cases, public concern has been warranted due to the limited amount of data on the fate and transport of pathogens after land application. Information is needed on indirect routes of human exposure, such as contact with bioaerosols at some distance from land application sites and consumption of ground water beneath sites. Therefore, over the past several years, the University of Arizona's National Science Foundation Water Quality Center has sponsored numerous studies on major biological issues related to land application of biosolids. In this review, we provide an evaluation of the relative risks of each hazard to human health based on a summary of the University of Arizona studies and other relevant studies.

Sustainability of land application has also been questioned based on the long-term effects of continuous biosolid applications on soil physical, microbial, and chemical properties. To evaluate these issues we document a long term University of Arizona study on the effects of land application on soil properties. Based on this study and a review of the literature, we provide an assessment of the sustainability of land application of Class B biosolids.

Copyright © 2008 by the American Society of Agronomy, Crop Science Society of America, and Soil Science Society of America. All rights reserved. No part of this periodical may be reproduced or transmitted in any form or by any means, electronic or mechanical, including photocopying, recording, or any information storage and retrieval system, without permission in writing from the publisher.

Published in *J. Environ. Qual.* 37:5-58-5-67 (2008).

doi:10.2134/jeq2007.0321

Received 15 June 2007.

*Corresponding author (ipepper@ag.arizona.edu).

© ASA, CSSA, SSSA

677 S. Segoe Rd., Madison, WI 53711 USA

I.L. Pepper, The Univ. of Arizona, Environmental Research Laboratory, 2601 E. Airport Drive, Tucson, AZ 85706; H. Zerzghi and C.P. Gerba, The Univ. of Arizona, Department of Soil, Water and Environmental Science, 1177 E. Fourth Street, Shantz Building, Room 429, Tucson, AZ 85721; J.P. Brooks, USDA ARS, Waste Management and Forage Research Unit, P.O. Box 5267, 810 Hwy 12 East, Mississippi State, MS, 39762.

Potential Biological Hazards Associated with Land Application of Class B Biosolids

Occurrence of Known Human Pathogens in Class B Biosolids

The known principal pathogens of concern in Class B biosolids are shown in Table 1. Pathogens are placed in the following categories: bacteria, viruses, protozoa, and helminth worms. The presence of *Salmonella*, enteric viruses, and helminth ova are used to distinguish Class A from Class B biosolids (USEPA, 1994). Class B biosolids may contain some or all of these known pathogens, and, because of this, restrictions meant to preclude initial human contact with biosolids are placed on land application sites. Essentially, site restrictions are implemented to allow for die-off of human pathogens to nondetectable levels. Despite this, public concern has focused on indirect routes of exposure such as bioaerosols to offsite locations or the leaching of pathogens into ground water. In addition, concerns have periodically arisen about the presence of specific or emerging human pathogens that could be present in biosolids. An example of this is the *Staphylococcus aureus* controversy of 2002.

Incidence of *S. aureus* in Sewage and Biosolids

S. aureus is the agent of a wide variety of skin and wound infections in humans and is commonly found in the nares and skin of warm blooded animals. Although there was no scientific documentation of *S. aureus* transmittal from wastewater or biosolids, claims were made that land-applied biosolids were the source of infections in residents living close to the application site (Lewis and Gattie, 2002). Staphylococci have been reported in aerosols from wastewater treatment plants using aeration treatments (Brandi et al., 2000). However, it was not confirmed that these were in fact *S. aureus*. Raw sewage was shown to be a definitive source of *S. aureus* in a more recent University of Arizona study, but it was not detected in Class A or B biosolids (Table 2), suggesting that the organism did not survive biosolid treatment (Rusin et al., 2003). In addition, the organism was never detected in bioaerosols resulting from land application sites. Based on these data, it was concluded that biosolids were not a significant source of *S. aureus* exposure (Rusin et al., 2003).

Offsite Exposure of Bioaerosols to Residents Close to Land Application Sites

Closely related to the *S. aureus* issue was the question of bioaerosols. Specifically, there was concern regarding the potential for movement of aerosolized pathogens away from a land application site to local residential communities. The risk of infection to residents living close to land application sites was identified as an area where data were lacking in the National Research Council report on land application (NRC, 2002). A limited number of studies have been conducted on the generation of bioaerosols from biosolids land application. Notably, Sorber et al. (1984) concluded that little or no risk was associated with the land application of liquid biosolids based on the lack of pathogenic viral presence in large volumes of sampled air. Other studies have focused on large piles of biosolids which were unloaded by trucks onsite and subsequently

Table 1. Known principal pathogens of concern in Class B biosolids in the U.S.

Bacteria	Protozoa
<i>Salmonella</i> sp.	<i>Cryptosporidium</i>
<i>Shigella</i> sp.	
<i>Yersinia</i>	<i>Giardia lamblia</i>
<i>Vibrio cholerae</i>	
<i>Campylobacter jejuni</i>	<i>Toxoplasma gondii</i>
<i>Escherichia coli</i>	
Enteric viruses	Helminth worms
Hepatitis A virus	<i>Ascaris lumbricoides</i>
Adenovirus	<i>Ascaris suum</i>
Norovirus	<i>Trichuris trichiura</i>
Sapporovirus	<i>Toxocara canis</i>
Rotavirus	<i>Taenia saginata</i>
Enterovirus	<i>Taenia solium</i>
Poliovirus	<i>Necator americanus</i>
Coxsackievirus	<i>Hymenolepisnana</i>
Echovirus	
Enterovirus 68–91	
Reovirus	
Astrovirus	
Hepatitis E virus	
Picobirnavirus	

loaded with front-end loaders into biosolid spreaders or hoppers (Pillai et al., 1996; Dowd et al., 2000). Loading events proved to be sources of increased concentration of fecal microbial indicators, such as H₂S producing bacteria and *Clostridium* spp. No risk analyses were conducted in the former study although the investigators concluded that the microbial indicator concentrations were below levels that could be construed as a risk to public health. The latter study conducted microbial risk analyses based on the use of complex transport models first proposed for the transport of chemical aerosols (Pasquill, 1961). Through the use of these models, aerosol concentrations could effectively be predicted at downwind distances from both point (biosolids pile) and area sources (a biosolids applied field; Dowd et al., 2000). Because of the lack of data on bioaerosols, the University of Arizona undertook a large national study to evaluate community risks from aerosols derived from land application sites (Brooks et al., 2005a, 2005b; Tanner et al., 2005). Risks of infection from *Salmonella* and Coxsackie virus A21 were determined at various distances from land-applied biosolids. An example of the data generated is illustrated in Table 3, which shows the annual community risk of infection from aerosolized *Salmonella* from land application at a distance of 30 m from the

Table 2. Incidence of *Staphylococcus aureus* in sewage, biosolids, and bioaerosols.†

Sample	No. of samples	Incidence of <i>S.aureus</i>
Raw sewage	3	1/3 positive
Undigested primary sewage	2	2/2 positive
Class B biosolids—anaerobic mesophilic	6	All negative
Class B biosolids—aerobic mesophilic	1	Negative
Class B biosolids—aerobic mesophilic, lime	9	All negative
Class A biosolids—aerobic thermophilic	1	Negative
Class A biosolids—anaerobic thermophilic	1	Negative
Class A biosolids—heat dried pellets	4	All negative
Class A composted biosolids	1	Negative
Bioaerosols from land application sites	27	All negative

† Source: Adapted from Rusin et al. (2003).

Table 3. Annual community risk of infection from aerosolized *Salmonella* during land application of Class B biosolid† (modified from Brooks et al., 2005b).

Annual risk					
No. of <i>Salmonella</i>					
500 <i>Salmonella</i> g ⁻¹ biosolids		50 <i>Salmonella</i> g ⁻¹ biosolids		5 <i>Salmonella</i> g ⁻¹ biosolids	
Exposure time					
1 h	8 h	1 h	8 h	1 h	8 h
4.5×10^{-8}	3.6×10^{-7}	4.5×10^{-9}	3.6×10^{-8}	4.5×10^{-10}	5.6×10^{-9}

† Risk of infection based on 6 d yr⁻¹ of land application at a distance of 30.5 m from the application site during loading conditions.

site. Risks were determined using the β -Poisson infectivity model for ingestion of nontyphoid *Salmonella* spp. (Brooks et al., 2005a). Risks of infection for Coxsackie virus A21 were also low. The most likely explanation for risks being so low is that for community risk, fate and transport of pathogens are important factors, allowing for dilution and natural attenuation of pathogens due to environmental factors such as dessication and ultraviolet light. In addition, it was postulated that organisms were bound to soil and/or biosolid particles with limited subsequent transport. Occupational risks

to biosolid workers were also evaluated and were found to be low, though still higher than community risks. The higher risks are due to enhanced duration of exposure and proximity to the land application site.

In a more recent study, the diversity of aerosolized bacteria during land application of Class B biosolids was determined (Brooks et al., 2007a). Specifically, sequence analyses of clones obtained from community DNA extractions were evaluated as to their source. The bacterial diversity of clone sequences was obtained from the following samples: (i) aerosols resulting from tractor operations on an agricultural field before land application (aerosol-soil); (ii) aerosols obtained during land application (aerosol-biosolids); (iii) Class B biosolids; and (iv) control soil (no biosolids). The most likely predominant source of aerosols obtained during land application of Class B biosolids can be determined from Fig. 1. Interpretation of the Venn diagram shows that the majority of bacterial aerosols obtained during land application of biosolids appear to be associated with the onsite soil, not the biosolids.

Recent work has concentrated on determining the source of bioaerosols from land application sites, utilizing methods devel-

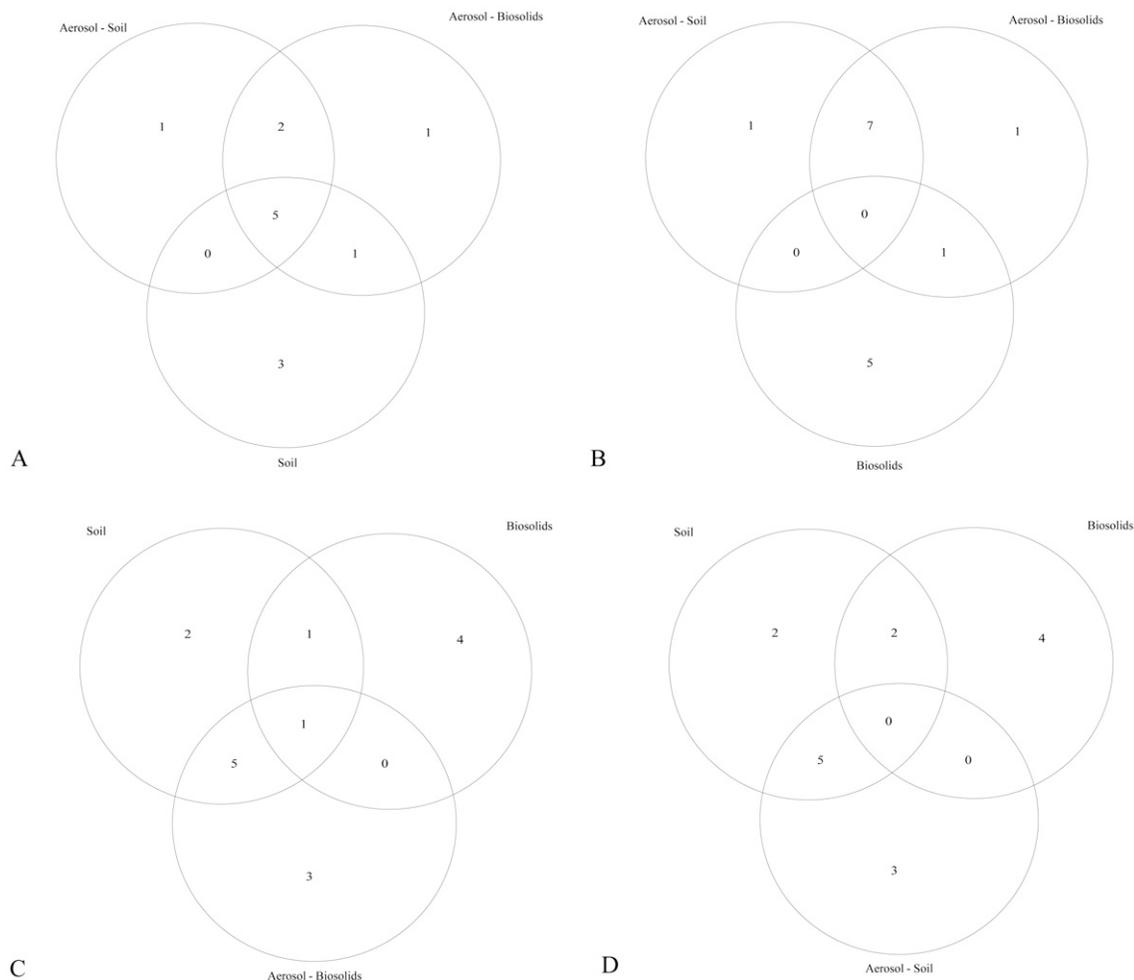


Fig. 1. (A) Number of shared orders associated with unique aerosolized clone isolates detected from downwind of aerosolized soil without biosolids, from downwind of biosolids land application operations when compared to onsite soil; (B) when compared to Class B biosolids; (C) Number of shared orders associated with unique clone isolates detected from onsite soil, Class B biosolids when compared to aerosols collected during biosolids land application operations; (D) when compared to aerosolized soil without biosolids. (Source: Brooks et al., 2007a).

oped for source-tracking fecal contamination in watersheds (Paez-Rubio et al., 2006; Baertsch et al., 2007; Paez-Rubio et al., 2007). Microbial source tracking (MST) methods were utilized, in which the authors used bacterial DNA from the class Chloroflexi and the phylum Euryarchaeota, which are typically present in high concentrations in biosolids (Baertsch et al., 2007). These MST methods consisted of screening concentrated aerosol samples for the presence of these organisms using polymerase chain reaction (PCR). In addition, the investigators also used innovative approaches to indirectly report bioaerosol concentrations associated with Class B biosolids sites in the desert southwestern United States (Paez-Rubio et al., 2006; Paez-Rubio et al., 2007). These approaches involved the use of particulate matter (PM) measurements along with bulk biosolid chemical and biological measurements. The biological and chemical compositions of downwind aerosols were estimated based on PM₁₀ and bulk biosolid measurements. Through these three studies it was determined that biosolids- and soil-derived particulate matter do in fact aerosolize during disking operations, often to concentrations greater than 1.5 mg m⁻³. The investigators estimate that biosolids-derived particulate matter make up approximately less than 1/1000th of this concentration, particularly during disking operations in which soil is the dominant particle. Overall this research confirmed that the majority of aerosols obtained during land application arise from soil sources rather than biosolids.

Based on these studies, we conclude that the community risk of human infection from pathogenic bioaerosols emanating from biosolids is low. However, another concern related to bioaerosols has been endotoxin, which can also be aerosolized. Endotoxin, lipopolysaccharide derived from the cell wall of Gram-negative bacteria, is a highly immunogenic class of molecules that, when introduced into the bloodstream or airways, can cause a broad range of health effects, such as fever, asthma, and shock (hence the suffix *toxin*; Bradley, 1979; Olenchock, 2001; Michel, 2003). Lipopolysaccharide is present throughout the environment, as Gram-negative bacteria continuously release lipopolysaccharide during both cell decay and active cell growth. Most surfaces contain some traces of endotoxin due to dust-associated endotoxin, and, therefore, most human populations come into contact with some endotoxin (Gereda et al., 2001; El Sharif et al., 2004). Although endotoxin is present in everyday environments, it is primarily of concern as an aerosol, since most human endotoxin ailments are pulmonary associated.

Exposure to aerosolized endotoxin has been studied with regard to occupational exposures from cotton dust, composting plants, and feed houses (Clark et al., 1983; Rylander et al., 1983; Castellan et al., 1987; Smid et al., 1992; Epstein, 1994; Donham et al., 2000). Exposures to levels of endotoxin as low as 0.2 endotoxin unit (EU) m⁻³ derived from poultry dust have been found to cause acute pulmonary ailments, such as decreases in forced expiratory volume (Donham et al., 2000). Chronic conditions such as asthma and bronchitis have been linked to daily exposures to endotoxin from cotton dust as low as 10 EU m⁻³ (Olenchock, 2001).

Past studies evaluating environmental exposures to endotoxin have used methods such as membrane-trapping of aerosolized endotoxin. Duchaine et al. (2001) compared two methods of collecting aerosolized endotoxin: traditional membrane trapping and

Table 4. Aerosolized endotoxin concentrations detected downwind of various operations (modified from Brooks et al., 2006).

Source of sample	No. of samples collected	Endotoxin concentration		
		Range	Avg	Median
Ambient background	12	2–4	2.6	2.5
Biosolids- loading	39	6–1808	344	92
Biosolids- land application	24	5–143	34	6
Wastewater treatment plant- aeration basin	6	294–891	627	639
Tractor on field- no biosolids	5	284–659	470	491

† EU = endotoxin units.

collection via impingement. Their results suggest that impingement may result in higher percent recoveries and greater precision. This same study focused on aerosolized endotoxin exposure in occupational settings, specifically swine barns and sawmills. Swine barns were found to contain mean concentrations of endotoxin 10 times greater than that of sawmills (4385 and 740 EU m⁻³, respectively). Endotoxin concentration ranged from a minimum of 208 to 17063 EU m⁻³ for sawmills and from 2026 to 11297 EU m⁻³ for swine barns, as collected by impingement sampling.

To determine the extent of endotoxin attributable to land application, aerosol samples were recently collected during land application of biosolids, during tractor operations on an agricultural field (no biosolids), and from an aeration basin located within an open-air wastewater treatment plant (Brooks et al., 2006). Data in Table 4 show that the greatest source of endotoxin was a wastewater treatment plant aeration basin, although differences were not significant. In addition, an evaluation of the agricultural operations shows that, on average, more endotoxin is aerosolized during tractor operations without biosolids than with biosolids. Therefore, the majority of endotoxin may in fact originate in the soil. Work conducted by Paez-Rubio et al. (2006, 2007) corroborated these findings. These data also agree with the Brooks et al. (2007a) study, which showed that soil was the major source of aerosolized bacteria.

Offsite Exposure to Pathogens via Ground Water Contamination

The transport and fate of viruses in porous media have received significant attention (Goyal and Gerba, 1979; Gerba et al., 1981; Bales et al., 1991, 1993, 1997; Jin et al., 1997; Dowd et al., 1998). Such studies have focused primarily on the adsorption, transport, and fate of viruses in aqueous (e.g., sewage effluent) systems. Conversely, minimal information with regard to the transport of indigenous coliphage associated with land application of biosolids is available.

The potential for the contamination of ground water with pathogens due to land application of biosolids is dependent on transport through soil and the vadose zone. Such transport is affected by a complex array of abiotic and biotic factors, including adhesion processes, filtration effects, physiological state of the cells, soil characteristics, water flow rates, predation, and intrinsic mobility of the organism (Newby et al., 2000). Recent work at the University of Arizona showed that viruses are embedded and/or adsorbed to biosolids, which likely influences the potential for

Table 5. Percentage of heterotrophic plate count (HPC) bacteria exhibiting antibiotic resistance from soil collected at a land application site (modified from Brooks et al., 2007b).

Time (days)	% Resistant†			
	Ampicillin	Cephalothin	Ciprofloxacin	Tetracycline
One day before land application	12.1	12.4	1.6	1.1
0	5.3	7.9	0.9	1.4
7	11.5	11.5	2.8	2.0
14	5.4	6.1	1.5	0.6
30	14.6	13.6	4.8	3.1
60	8.7	10.7	1.9	1.6
90	8.2	8.4	2.1	1.8
120	8.4	11.0	2.2	2.0
150	9.1	11.1	2.0	1.8
180	5.3	6.6	2.7	1.3
450	6.7	10.5	2.3	1.9
Mean value post biosolid land application (7–450 d)	8.7	9.9	2.3	1.9

† Antibiotic concentrations of cultured plates: ampicillin = 32 µg mL⁻¹; cephalothin = 32 µg mL⁻¹; ciprofloxacin = 4 µg mL⁻¹; and tetracycline = 16 µg mL⁻¹.

release and subsequent transport of the virus through soil under saturated conditions (Chetochine et al., 2006). Overall, <8% of indigenous coliphage initially present in Class B biosolids were leached out of the biosolids/soil matrix. Such binding reduces the mobility of viruses and limits the potential for significant leaching. An additional study also showed that dissolved organic carbon had no influence on viral transport (Cheng et al., 2007). In agreement with this, Pancorbo et al. (1981) seeded anaerobically digested sewage sludge with poliovirus type 1 to a concentration of 10⁶ plaque forming units (PFU) mL⁻¹. They recovered 60% by use of a serial extraction protocol suitable for poliovirus. Their findings indicated that the seeded polioviruses were adsorbed to the solid component of the sewage. Wellings et al. (1976) and Stagg et al. (1978) both reported that indigenous viruses were solid-associated in wastewater-activated sludge. Stagg et al. demonstrated that 85% of the indigenous coliphage was adsorbed to solids present in sewage effluent. Bales et al. (1993) also reported that organic matter inhibited transport of phage and human viruses due to hydrophobic interactions. In contrast to these studies, other researchers have demonstrated enhanced transport of viruses in the presence of organic matter (Pieper et al., 1997; Jin et al., 2000; Jin and Flury, 2002; Bradford et al., 2006b). This enhanced transport has been attributed to the blocking of favorable attachment sites by organic matter (Guber et al., 2005a). Dissolved organic matter from manure suspensions has also been shown to enhance transport of bacteria in soil (Guber et al., 2005b). Filling of straining sites has also been used to explain this enhanced transport (Bradford et al., 2006a). The different results obtained by different researchers are most likely due to the different types and concentrations of organic matter used in these studies. In the University of Arizona study (Chetochine et al., 2006), phage were shown to be tightly sorbed to biosolids such that 58 sequential extractions of the same biosolid sample were required to remove all phage from the biosolids. Thus, adsorbed and/or embedded viruses are less likely to be transported. Based on this, ground water contamination from land application of biosolids does not appear likely—other than in areas where karst soils predominate with the potential for preferential flow.

Antibiotic-Resistant Bacteria

The antibiotic era began following Alexander Fleming's discovery of penicillin in 1929. Since the introduction of antibiotics, overuse has been an issue, and over-prescription of first-generation antibiotics has led to many resistant bacterial strains (Monroe and Polk, 2000; Lieberman, 2003). Human bacterial pathogens such as *Salmonella*, *Shigella*, and *Campylobacter* can potentially all be present in biosolids. This may be cause for concern due to the potential for gene transfer, which could include antibiotic resistance (Rensing et al., 2002; Dzidic and Bedekovic 2003). Of these, pathogenic antibiotic resistant bacteria (ARB) strains may be of most concern as these organisms are already resistant to many commonly prescribed antibiotics (Low, 2001; Marshall et al., 2004). Non-pathogenic ARB can potentially transfer their resistance genes horizontally to pathogenic strains through antibiotic resistant plasmids, either in the environment or human host (Rensing et al., 2002; Dzidic and Bedekovic, 2003; Salyers et al., 2004). Therefore, when soil, water, or food that has been in contact with biosolids is consumed, these ARB may be present, and the subsequent transfer of resistance genes occurring within the host is a possibility. To date, however, this has not been demonstrated.

The influence of land application of biosolids on the incidence of ARB within soil was recently investigated (Brooks et al., 2007b). Table 5 illustrates the influence of land application of Class B biosolids on the incidence of soil bacterial resistance to four antibiotics: ampicillin, cephalothin, ciprofloxacin, and tetracycline. Data from soil samples collected before land application, and up to 450 d following land application show that the influence of biosolids on the incidence of soil-borne antibiotic resistant bacteria was negligible.

Risk of Infection from *Salmonella* Regrowth in Biosolids

Regrowth of *Salmonella* in biosolids being stored after having left the wastewater treatment plant is one of the more recent concerns with respect to land application of biosolids. Many laboratory studies have been conducted, focusing on survival and potential growth of inoculated organisms in sterile and nonsterile biosolids and compost. Regrowth of inoculated salmonellae in sterile biosolids is commonly documented, but few studies have documented survival and regrowth of indigenous pathogens in biosolids after levels have decreased below levels of detection. Due to limited available data regarding regrowth of salmonellae in biosolids used for land application, there are conflicting views on this topic. Some studies have shown that regrowth does occur, while other studies have shown that regrowth does not occur. These differences are most likely due to whether indigenous salmonellae are studied or whether biosolids are reseeded with salmonellae. Recently the regrowth potential of both Class A and Class B biosolids and biosolid amended soil was evaluated at the University of Arizona (Zaleski et al., 2005a; Castro del Campo et al., 2007).

Regrowth of *Salmonella* does not normally occur in Class B biosolids (Zaleski et al., 2005a). However, during solar drying of Class B liquid biosolids in concrete lined drying beds, regrowth

was observed. During solar drying, the initially Class B biosolids material (both aerobically and anaerobically) attained Class A status with respect to *Salmonella* levels (<3 most probable number [MPN] per 4 g). However, following rainfall events, the biosolids became saturated and, hence, anaerobic, allowing regrowth of *Salmonella* to 10^5 MPN g^{-1} (Zaleski et al., 2005b). For Class A biosolids, regrowth also occurred when biosolids were maintained under saturated anaerobic conditions (Castro-del Campo et al., 2007). This is consistent with other cited studies. For example Burge et al. (1987) observed that growth of *Salmonella* in Class B biosolids required moisture content greater than 20%. This agrees with Thomason et al. (1975), who also detected *Salmonella* within wet environments. No regrowth occurred from Class A or B biosolids when the material was added to soil, regardless of whether samples were saturated or not (Zaleski et al., 2005b). The risks of infection from *Salmonella* from direct contact with, or aerosolization from, land-applied Class A biosolids in which regrowth had occurred or land-applied Class B biosolids are shown in Table 6. Risks from land-applied Class B biosolids were low regardless of whether the route of exposure was from the ingestion of *Salmonella* following direct exposure or from ingestion following inhalation of an aerosol. Note that *Salmonella* infections due to inhalation have not been documented at this time. In contrast, risks from contact with Class A biosolids following regrowth in the initial biosolids were significant. Therefore, care must be taken to prevent regrowth in Class A biosolids before land application. Regrowth can be prevented by covering biosolids and precluding saturated anaerobic conditions (Gerba et al., 2007).

Long-Term Effects of Land Application of Liquid Class B Biosolids on Soil Properties

Sustainability of any process implies that the process can be maintained indefinitely. Further, sustainability necessitates addressing the needs of the present without compromising the ability of future generations to meet their needs. Within this context, the sustainability of land application of Class B biosolids needs to be evaluated over long time periods, and the influence of repeated annual applications of biosolids on soil properties must be documented. Fundamentally, soil quality is maintained through the integration of soil physical, microbial, and chemical properties.

Influence on Soil Physical Properties

Soil physical properties are controlled by soil texture, which is intrinsic to any given soil, and soil structure, which is highly variable. Long-term additions of biosolids have been shown in several studies to improve the physical properties of the amended soil. Aggelides and Landra (2000) documented improved physical properties including saturated and unsaturated hydraulic conductivity, water retention capacity, soil resistance to penetration, bulk density, total porosity, pore size distribution, aggregation, and aggregate instability. This study was conducted in a semiarid climate and improvements were greater in a loamy soil than in a clay soil. Similarly, even 3 yr of land application of biosolids improved the physical properties of a clay loam soil (Tsadilas et al., 2005). This field study was conducted under Mediterranean conditions in

Table 6. Risk of infection from *Salmonella* from direct contact with, or aerosolization from, land-applied Class A (following regrowth) or Class B land-applied biosolids (modified from Gerba et al., 2007).

Class of biosolids land applied	No. of <i>Salmonella</i> /gm	Route of exposure	Risk of infection
B	105	Ingestion of 50 mg via direct contact	6×10^{-6}
B	105	Ingestion of 480 mg via direct contact	6×10^{-5}
B	500	Ingestion of 50% of aerosol† inhaled	1×10^{-4}
A	10^6	Ingestion of 50 mg via direct contact	5×10^{-3}
A	10^6	Ingestion of 480 mg via direct contact	3×10^{-1}
A	10^6	Ingestion of 50% ¹ of aerosol inhaled	1×10^{-2}

† 8 h exposure per day for 6 d annually at 30 m distance from the site.

central Greece and documented enhanced water retention capacity, available water, and infiltration rates. The improvement of physical properties is most likely due to the increased soil organic matter content following land application. Organic matter content has been positively correlated with water retention and infiltration rate, and negatively correlated with bulk density and aggregate instability index (Tsadilas et al., 2005). Although the increased soil organic matter content that occurs following an application is always likely to improve soil physical properties, care must be taken to avoid compaction that can occur due to increased traffic operations. Stamatidis et al. (1999) documented soil compaction and decreased water infiltration rates following land application of liquid municipal sewage sludge in eastern Nebraska. However, if land application is carefully managed (i.e., preventing traffic over saturated soils), then land application is likely to be sustainable with respect to soil physical properties.

Influence on Soil Microbial Properties

The influence of long-term land application on the soil microbial community and soil chemical properties was evaluated in a field study conducted from 1986 to 2005 by the University of Arizona (Zerzghi, 2008). As one of the longest studies of its kind, this 20-yr project provides a basis for evaluating the sustainability of land application. This replicated field-plot study had 4 treatments: (i) control (no amendment); (ii) inorganic fertilizer control; (iii) biosolids at an agronomic rate ($1 \times$) based on the N requirements for the growth of cotton (160 kg N ha^{-1}); and (iv) biosolids at a $3 \times$ rate. The land-applied biosolids contained 8% solids. Twenty annual land applications were administered from 1986 to 2005. All plots were utilized for the growth of cotton and were furrow irrigated as necessary for optimum plant growth. Approximately 100 cm of irrigation water was supplied per growing season, since the consumptive water use requirements for cotton are 92 cm per season. This is approximately twice the evapotranspiration requirements for cotton, allowing for leaching of soluble salts through the soil profile. The influence of land application on the soil microbial community was evaluated in terms of microbial numbers, microbial activity, and microbial diversity. The survival of indicator and pathogenic organisms was also determined. Finally, note that some parameters were evaluated at various points throughout the 20-yr study (Artiola and Pepper, 1992; Brendecke et al., 1993).

Land application of Class B biosolids resulted in numbers of heterotrophic bacteria, fungi, and actinomycetes that were similar to numbers found in control plots. This is not unexpected since the surface soil samples for microbial analyses were collected in

Table 7. Influence of 20 yr of land application of liquid Class B biosolids on soil chemical properties. Adapted from Zerzghi (2008).

Soil chemical parameter	Influence of land application
pH	None
Electrical conductivity (EC)	None
% CaCO ₃	Values increased with rate of biosolid addition and soil depth
Total N	Higher in biosolid plots
Total organic carbon	Higher in biosolid plots
Total P	Higher in biosolid plots
Available P	Higher in biosolid plots
Total Metals: Cu; Cd; Zn	Higher in biosolid plots
Total Metals: Pb; Ni	No discernable effect
DTPA† Metals: Cu; Zn; Pb; Ni	Higher in biosolid plots
DTPA Cd	No discernable effect
Total Metals: As; Hg; Mo; Se; Cr and B	No discernable effect
Polybrominated diphenyl ethers (PBDEs)	Higher in biosolid plots

† DTPA = diethylene triamine penta acetic acid.

December 2005, 9 mo after the last application of biosolids in March 2005. During this interval, a crop of cotton was grown and harvested, and all available microbial substrate would have been utilized. The lack of adverse effects of 20 yr of land application of biosolids on soil microbial numbers agrees with analyses within this study that were conducted in 1990 after four annual land applications (Brendecke et al., 1993). In these analyses, 4 yr of land application had no significant effect on numbers of soil bacteria, fungi, or actinomycetes. In addition, soil respiration rates (as measured by CO₂ evolution) were similar in land-applied and control plots. In a recent British study, 3 yr of land application at three different sites likewise resulted in no observed effect on soil respiration rates or biomass carbon concentrations (Gibbs et al., 2006a). In addition, in this study, numbers of indigenous clover *Rhizobium* were also unaffected by biosolid additions. After 20 yr of land application in the University of Arizona study, total coliform and fecal coliform counts did not exceed 3 MPN g⁻¹ in plots that received biosolids. Control and fertilizer control plots did not contain detectable indicators or coliphage. *Salmonella* and enteroviruses were not detected in any plots.

These data agree with other studies on the survival of indicators and pathogens introduced into soil via land application of biosolids. Specifically, limited survival of *E. coli* and other enteric organisms in biosolids-amended agricultural soil was documented in laboratory and field studies (Lang and Smith, 2007; Lang et al., 2007; Lang et al., 2003; Pepper et al., 1993). Lang and Smith (2007) concluded that the indigenous soil biota were involved in pathogen reduction processes in biosolid-amended soil. Survival of enteroviruses in biosolid-amended soils was also evaluated earlier in the University of Arizona study. Specifically the duration of survival of poliovirus type 1 was less than 7 d in the summer months, and 7–14 d during winter months (Straub et al., 1993). Thus, it is not surprising that viruses were not detected in the University of Arizona study 9 mo after the last land application.

Microbial activity in the University of Arizona 20-yr land application study was assessed by studies on the common microbial transformations, nitrification and sulfur oxidation, where rates of both microbial transformations increased in soils from the biosolid amended plots and increased with increased rate of biosolids application. Dehydrogenase activity also increased with increased

biosolid amendment. This is in contrast to trends noted after 4 yr of land application, when dehydrogenase activity was unaffected by biosolid additions (Brendecke et al., 1993). After 20 yr of land application, bacterial diversity in all plots was evaluated through cloning and sequence analysis of bacterial 16S rRNA (Zerzghi, 2008). Here, data showed that the known number of identifiable species increased in the high-rate biosolid plots when compared to control (no biosolid) plots.

In summary, the soil microbial community did not appear to be adversely affected by 20 yr of land application. In fact, land application appeared to have been beneficial as evidenced by increased microbial diversity and enhanced microbial activity. In addition, no known pathogens were detected in soils sampled 9 mo after the last biosolid application.

Influence on Soil Chemical Properties

At the termination of the 20-yr land application study, soil chemical properties were also evaluated in all plots. Specifically, soil core samples were collected at depths of 0–150 cm in 30-cm intervals. As in the case for surface soil samples collected for microbial analyses, samples were taken in December 2005, 9 mo after the 20th land application in March 2005. These soil core subsurface samples allowed for an evaluation not only of the influence of long-term land application of biosolids on soil chemical properties, but also the influence of land application on various parameters as a function of soil depth. Analyzed soil chemicals clustered into three main groups: soil macro elements (N, P, and C), heavy metals, and endocrine disruptors.

Soil Macro Elements

Soil nitrate concentrations in both biosolid amended plots and plots that received inorganic fertilizers for 20 yr were significantly higher than control (no amendment) plots when averaged over all soil depths (0–150 cm). Total N increased in biosolid-amended soil (Table 7). Nitrate values in both biosolid and fertilizer treated plots exceeded 10 ppm NO₃-N at most soil depths down to 150 cm. These data indicate the potential for nitrate pollution of ground water regardless of whether biosolids or inorganic fertilizers are applied to the soil. Land application of biosolids also significantly increased total and available soil phosphate concentrations, particularly in the surface (0–30 cm) soil. These data are not unexpected since several other studies have documented phosphate increases following land application (Mantovi et al., 2005). Increases in soil phosphate concentrations of biosolid amended soil were already evident in soil samples collected from the same study after 4 yr of land application (Brendecke et al., 1993). Elliott and O'Connor (2007, p. 1319) recently stated that “phosphorus is at the forefront of biosolids-related issues that may adversely affect the long term sustainability of land-based recycling programs in the U.S.” Such concern is based on the potential for water quality deterioration that can occur in surface waters due to eutrophication following phosphate accumulations in surface water runoff. However, these issues are more important in the eastern United States since surface waters are rare in the arid southwest. That notwithstanding, P management will continue to be important for sustainable biosolids recycling in the United States.

Total organic C increased significantly in biosolid-amended soil after 20 yr of land application in the University of Arizona study. This is in contrast to analyses of the same plots following five land applications. Data from samples collected in 1990 showed no differences in soil total organic C in control versus biosolid-amended soils (Artiola and Pepper, 1992). However, dissolved organic C increases were detected in the 1990 biosolid-amended soil samples. These data illustrate how difficult it is to increase soil organic matter in soils of the arid southwestern United States—a result of high mineralization rates (Artiola and Pepper, 1992). But it is important to note that even modest increases in total organic C are important to fertility in soils that are traditionally low in soil organic matter (0.5–1%; Fuller, 1991). Land application of biosolids has also been shown to increase soil organic C in other studies (Gibbs et al., 2006b; Mantovi et al., 2005).

No increases in soil salinity were observed following 20 yr of land application. This is not surprising given that irrigation rates were in excess of consumptive water use rates for cotton, resulting in salts being leached through the soil profile.

Soil Heavy Metals

The biosolids applied in the University of Arizona 20-yr land application study contained relatively low levels of heavy metals (Table 8). After 20 annual land applications, significant but modest increases in available concentrations were detected for some metals, including Cu, Cd, Zn, and Ni (Table 7). However, biosolid-amended soil concentrations of available metals were low and not hazardous. In addition, the metal concentrations found within biosolids have decreased over the past 20 yr due to improved point-source controls. Finally, metal concentrations attenuated rapidly with increasing soil depth and were generally similar to values found in control soils at a depth of 150 cm. Overall, the potential for metal contamination of soil or surface waters from land application of biosolids has decreased. Recently, Tian et al. (2006) concluded that application of biosolids for land reclamation at high loading rates from 1972 to 2002 only impacted surface water quality before the promulgation of the 40 Code of Federal Regulations Part 503 regulations. After the promulgation, metal impacts on surface water quality were minimal. In addition to reduced concentrations of metals in biosolids, new understanding of trace element chemistry in biosolid-amended soil has shown that following termination of land application, available metal concentrations essentially remain constant, or even decrease, provided the soil pH remains constant (Basta et al., 2005). Such decreases are thought to be caused by sorption to inorganic oxide surfaces or very recalcitrant organics of non-biosolid origin present in soil (Basta et al., 2005).

Endocrine Disruptors

Endocrine disrupting compounds (EDCs) are chemicals that interfere with endocrine glands or their hormones. Polybrominated diphenyl ethers (PBDEs) are compounds utilized as flame retardants for everyday household items including carpets and cushions. Polybrominated diphenyl ethers are a known class of EDCs that are typically present at ppm levels in municipal biosolids produced in the United States. Data on the fate of PBDEs

Table 8. Representative heavy metals in land-applied biosolids in the University of Arizona study.

Metal	Range (mg kg ⁻¹ dry biosolids)
Zn	800–1590
Cu	568–957
Pb	89–221
Ni	26–51
Cr	32–95
Cd	7–15
Ag	3–60

following land application are limited, though this was recently evaluated at the University of Arizona. Following 20 yr of land application, 50–70% of the (estimated) applied PBDE mass was accounted for in the upper - 60 cm of receiving soil, suggesting that PBDEs are conserved in soil over periods of decades or longer, most likely due to the hydrophobic nature of PBDEs (Quanrud et al., 2007). Due to sorption, biodegradation of the PBDEs would be unlikely. It is important to note that although the consequences of PBDE accumulation in soil are unknown, PBDE concentrations in household dust are similar to concentrations found in biosolids (Quanrud et al., 2007).

Biosolids are also known to contain other EDCs, including estradiol and estrone (Drewes and Shore, 2001), and can contain other synthetic EDCs present in pharmaceuticals and personal care products (Daughton and Ternes, 1999). Little is currently known, however, of the fate and transport of these compounds following land application. Lorenzen et al. (2006) showed that a wide range of EDCs, including 4-nonylphenol, ethynylestradiol, estradiol, and estrone, are rapidly degraded from biosolids or animal wastes following land application. Roberts et al. (2006) also showed that 4-nonylphenol was rapidly mineralized in soil. In addition, they found that plant uptake of nonylphenol was minimal. Finally, they concluded that “the spreading of nonylphenol contaminated waste to soil probably poses a very low environmental risk to freshwater ecosystems and human health” (Roberts et al., 2006, p. 1812). Based on these results, land application of biosolids is likely to be sustainable with respect to EDCs.

Summary: Sustainability of Land Application of Class B Liquid Biosolids

The conclusion we have reached, based on all of our land application studies over the past two decades and an in-depth review of other relevant land application studies, is that land application of Class B biosolids is sustainable. Specifically, the risks to human health posed by microbiological entities within biosolids have been shown to be low if current USEPA regulatory guidelines are followed. In addition, risks from indirect exposures, such as aerosolized pathogens or contaminated ground waters, appear to be particularly low. This is not to say that the risks are zero, but that the risks or concerns can be managed to safeguard human health and provide a sustainable environment. Of course, vigilance is always necessary—as new contaminants continue to emerge, their presence and fate in biosolids need to be assessed to ensure current practices and guidelines are protective. Long-term land application in the University of Arizona study showed enhanced microbial activity and diversity. Overall these are positive

beneficial effects, and no evidence of adverse toxicity effects was found on the soil microbial community with respect to numbers, activity, or diversity. Long-term land application also increased soil macronutrients, including C, N, and P. In addition, increases in available metal concentrations were modest, and no increase in soil salinity was observed. This lack of increased salinity was likely due to leaching of salts out of the root zone following row irrigations throughout each growing season. Since irrigation rates were in excess of consumptive water use rates for cotton, such leaching may ultimately impact ground water quality. To date, there does not appear to be any chemical entity likely to limit land application, with the possible exception of phosphate loadings in areas sensitive to surface water contamination.

Additionally, the University of Arizona study revealed no evidence of long-term persistence of enteric pathogens in the soil or migration of pathogens to ground water. Further, it has been well documented that EDCs such as 4-nonylphenol and estrone are rapidly degraded in soils. Other hydrophobic endocrines such as PBDEs are strongly sorbed to soil colloids and are relatively immobile in soil. Finally, it should be noted that the University of Arizona study was conducted in the arid southwest and, as such, results are site-specific. Application of these findings at other locations needs to be conducted carefully.

References

- Aggelides, S.M., and P.A. Landra. 2000. Effects of compost produced from town wastes and sewage sludge on the physical properties of a loamy and a clay soil. *Bioresour. Technol.* 71:253–259.
- Artiola, J.F., and I.L. Pepper. 1992. Long term influence of liquid sewage sludge on the organic carbon and nitrogen content of a furrow-irrigated desert soil. *Biol. Fertil. Soils* 14:30–36.
- Baertsch, C., T. Paez-Rubio, E. Viau, and J. Peccia. 2007. Source tracking aerosols released from land-applied class B biosolids during high-wind events. *Appl. Environ. Microbiol.* 73:4522–4531.
- Bales, R.C., S.R. Hinkle, T.W. Kroeger, K. Stocking, and C.P. Gerba. 1991. Bacteriophage adsorption during transport through porous media: Chemical perturbations and reversibility. *Environ. Sci. Technol.* 25:2088–2095.
- Bales, R.C., S. Li, K.M. Maguire, M.T. Yahya, and C.P. Gerba. 1993. MS-2 and poliovirus transport in porous media: Hydrophobic effects and chemical perturbations. *Water Resour. Res.* 29:957–963.
- Bales, R.C., S. Li, T.C.J. Yeh, M.E. Lenczewski, and C.P. Gerba. 1997. Bacteriophage and microsphere transport in saturated porous media: Force-gradient experiment at Borden, Ontario. *Water Resour. Res.* 33:639–648.
- Basta, N.T., J.A. Ryan, and R.L. Chaney. 2005. Trace element chemistry in residual-treated soil: Key concepts and metal bioavailability. *J. Environ. Qual.* 34:49–63.
- Bradford, S.A., J. Simunek, and S.L. Walker. 2006a. Transport and straining of *E. coli* 0157:H7 in saturated porous media. *Water Resour. Res.* 42, W12S12, doi:10.1029/2005WR4805.
- Bradford, S.A., Y.F. Tadassa, and Y. Jin. 2006b. Transport of coliphage in the presence and absence of manure suspensions. *J. Environ. Qual.* 35:1692–1701.
- Bradley, S.G. 1979. Cellular and molecular mechanisms of action of bacterial endotoxins. *Annu. Rev. Microbiol.* 33:67–94.
- Brandi, G., M. Sisti, and G. Amagliani. 2000. Evaluation of the environmental impact of microbial aerosols generated by wastewater treatment plants utilizing different aeration systems. *J. Appl. Microbiol.* 88:845–852.
- Brendecke, J.W., R.D. Axelson, and I.L. Pepper. 1993. Soil microbial activity as an indicator of soil fertility: Long-term effects of municipal sewage sludge on an arid soil. *Soil Biol. Biochem.* 25:751–758.
- Brooks, J.P., C.P. Gerba, and I.L. Pepper. 2007a. Diversity of aerosolized bacteria during land-application of biosolids. *J. Appl. Microbiol.* 103:1779–1790.
- Brooks, J.P., P.A. Rusin, S.L. Maxwell, C. Rensing, C. Gerba, and I.L. Pepper. 2007b. Occurrence of antibiotic-resistant bacteria and endotoxin associated with the land-application of biosolids. *Can. J. Microbiol.* 53:1–7.
- Brooks, J.P., B.D. Tanner, C.P. Gerba, C.N. Haas, and I.L. Pepper. 2005a. Estimation of bioaerosol risk of infection to residents adjacent to a land applied biosolids site using an empirically derived transport model. *J. Appl. Microbiol.* 98:397–405.
- Brooks, J.P., B.D. Tanner, C.P. Gerba, and I.L. Pepper. 2006. The measurement of aerosolized endotoxin from land-application of Class B biosolids in Southeast Arizona. *Can. J. Microbiol.* 52:150–156.
- Brooks, J.P., B.D. Tanner, K.L. Josephson, C.N. Haas, C.P. Gerba, and I.L. Pepper. 2005b. A national study on the residential impact of biological aerosols from the land-application of biosolids. *J. Appl. Microbiol.* 99:310–322.
- Burge, W.D., N.K. Enkiri, and D. Hussong. 1987. *Salmonella* regrowth in compost as influenced by substrate (*Salmonella* regrowth in compost). *Microbiol. Ecol.* 14:243–253.
- Castellan, R.M., S.A. Olenchok, K.B. Kinsley, and J.L. Hankinson. 1987. Inhaled endotoxin and decreased spirometric values: An exposure-response relation for cotton dust. *N. Engl. J. Med.* 317:605–610.
- Castro-del Campo, N., I.L. Pepper, and C.P. Gerba. 2007. Assessment of *Salmonella typhimurium* growth in Class A biosolids and soil/biosolid mixtures. *J. Res. Sci. Technol.* 4:83–88.
- Cheng, L., A.S. Chetochine, I.L. Pepper, and M.L. Brusseau. 2007. Influence of DOC on MS-2 bacteriophage transport in a sandy soil. *Water Air Soil Pollut.* 178:315–322.
- Chetochine, A., M.L. Brusseau, C.P. Gerba, and I.L. Pepper. 2006. Leaching of phage from Class B biosolids and potential transport through soil. *Appl. Environ. Microbiol.* 72:665–671.
- Clark, C.S., R. Rylander, and L. Larsson. 1983. Levels of Gram-negative bacteria, *Aspergillus fumigatus*, dust, and endotoxin at compost plants. *Appl. Environ. Microbiol.* 45:1501–1505.
- Daughton, C.G., and T.A. Ternes. 1999. Pharmaceuticals and personal care products in the environment: Agents of subtle change. *Environ. Health Perspect.* 107:907–938.
- Donham, K.J., D. Cumro, S.J. Reynolds, and J.A. Merchant. 2000. Dose-response relationships between occupational aerosol exposures and cross-shift declines of lung function in poultry workers: Recommendations for exposure limits. *J. Occup. Environ. Med.* 42:260–269.
- Dowd, S.E., S.D. Pillai, S. Wang, and M.Y. Corapcioglu. 1998. Delineating the specific influence of virus isoelectric point and size on virus adsorption and transport through sandy soils. *Appl. Environ. Microbiol.* 64:405–410.
- Dowd, S.E., C.P. Gerba, I.L. Pepper, and S.D. Pillai. 2000. Bioaerosol transport modeling and risk assessment in relation to biosolid placement. *J. Environ. Qual.* 29:343–348.
- Drewes, C.G., and L.A. Shore. 2001. Concerns about pharmaceuticals in water reuse, ground water recharge and animal waste. p. 206–228. *In* C.G. Daughton and T.L. Jones-Lepp (ed.) *Pharmaceuticals and personal care products in the environment: Scientific and regulatory issues*. American Chemical Society, Washington, DC.
- Duchaine, C., P.S. Thorne, A. Merizux, Y. Grimard, P. Whitten, and Y. Cormier. 2001. Comparison of endotoxin exposure assessment by bioaerosol impinger and filter-sampling methods. *Appl. Environ. Microbiol.* 67:2775–2780.
- Dzidic, S., and V. Bedekovic. 2003. Horizontal gene transfer-emerging multidrug resistance in hospital bacteria. *Acta Pharmacol. Sin.* 24:519–526.
- El Sharif, N., J. Douwes, P.H.M. Hoet, G. Doekes, and B. Nemery. 2004. Concentrations of domestic mite and pet allergens and endotoxin in Palestine. *Allergy* 59:623–631.
- Elliott, H.A., and G.A. O'Connor. 2007. Phosphorus management for sustainable biosolids recycling in the United States. *Soil Biol. Biochem.* 39:1318–1327.
- Epstein, E. 1994. Composting and bioaerosols. *BioCycle* 35:51–58.
- Fuller, W.H. 1991. Organic matter applications. p. 507–541. *In* J. Skujins (ed.) *Semiarid lands and deserts: Soil resource and reclamation*. Marcel Dekker, New York.
- Gerba, C.P., N. Castro-del Campo, J.P. Brooks, and I.L. Pepper. 2007. Exposure and risk assessment of *Salmonella* in recycled residuals. IWA 6th Conf. on Wastewater and Reclamation and Reuse for Sustainability, 9–12 Oct. 2007, Antwerp, Belgium.
- Gerba, C.P., S.M. Goyal, I. Cech, and G.F. Bogdan. 1981. Quantitative assessment of the adsorptive behavior of viruses to soils. *Environ. Sci. Technol.* 15:940–944.
- Gereda, J.E., M.D. Klinnert, M.R. Price, D.Y.M. Leung, and A.H. Liu. 2001. Metropolitan home living conditions associated with indoor endotoxin levels. *J. Allergy Clin. Immunol.* 107:790–796.

- Gibbs, P.A., B.J. Chambers, A.M. Chaudri, S.P. McGrath, and C.H. Carlton-Smith. 2006a. Initial results from long-term field studies at three sites on the effects of heavy metal-amended liquid sludges on soil microbial activity. *Soil Use Manage.* 22:180–187.
- Gibbs, P.A., B.J. Chambers, A.M. Chaudri, S.P. McGrath, C.H. Carlton-Smith, J.R. Bacon, C.D. Campbell, and M.N. Aitken. 2006b. Initial results from a long term multi-site field study of the effects on soil fertility and microbial activity of sludge cakes containing heavy metals. *Soil Use Manage.* 22:11–21.
- Goyal, S.M., and C.P. Gerba. 1979. Comparative adsorption of human enteroviruses simian rotavirus, and selected bacteriophages to soils. *Appl. Environ. Microbiol.* 38:241–247.
- Guber, A.K., D.R. Shelton, and Y.A. Pachepsky. 2005a. Transport and retention of manure-borne coliforms in undisturbed soil columns. *Vadose Zone J.* 4:828–837.
- Guber, A.K., D.R. Shelton, and Y.A. Pachepsky. 2005b. Effect of manure on *Escherichia coli* attachment to soil. *J. Environ. Qual.* 34:2086–2090.
- Jin, Y., and M. Flury. 2002. Fate and transport of viruses in porous media. *Adv. Agron.* 77:39–102.
- Jin, Y., E. Pratt, and M.V. Yates. 2000. Effect of mineral colloids on virus transport through saturated sand columns. *J. Environ. Qual.* 29:532–539.
- Jin, Y., M.V. Yates, S.S. Thompson, and W.A. Jury. 1997. Sorption of viruses during flow through saturated sand columns. *Environ. Sci. Technol.* 31:548–555.
- Lang, N.L., M.D. Bellett-Travers, and S.R. Smith. 2007. Field investigations on the survival of *Escherichia coli* and presence of other enteric microorganisms in biosolids-amended agricultural soil. *J. Appl. Microbiol.* 103:1868–1882.
- Lang, N.L., and S.R. Smith. 2007. Influence of soil type, moisture content and biosolids application on the fate of *Escherichia coli* in agricultural soil under controlled laboratory conditions. *J. Appl. Microbiol.* 103:2122–2131.
- Lang, N.L., S.R. Smith, D.M. Bellett-Travers, E.B. Pike, and C.I. Rowlands. 2003. Decay of *Escherichia coli* in soil following the application of biosolids to agricultural land. *Water Environ. Manage. J.* 17:23–28.
- Lewis, D.L., and D.F. Gattie. 2002. Pathogen risks from applying sewage sludge to land. *Environ. Sci. Technol.* 36:286A–293A.
- Lieberman, J.M. 2003. Appropriate antibiotic use and why it is important: The challenges of bacterial resistance. *Pediatr. Infect. Dis. J.* 22:1143–1151.
- Lorenzen, A., K. Burnison, M. Servos, and E. Topp. 2006. Persistence of endocrine disrupting chemicals in agricultural soils. *J. Environ. Eng. Sci.* 5:211–219.
- Low, D.E. 2001. Antimicrobial drug use and resistance among respiratory pathogens in the community. *Clin. Infect. Dis.* 33(Suppl. 3):S206–S213.
- Mantovi, P., G. Baldoni, and G. Toderi. 2005. Reuse of liquid dewatered and composted sewage sludge on agricultural land: Effects of long term application on soil and crop. *Water Res.* 39:289–296.
- Marshall, C., S. Wesselingh, M. McDonald, and D. Spelman. 2004. Control of endemic MRSA—what is the evidence? A personal view. *J. Hosp. Infect.* 56:253–268.
- Michel, O. 2003. Role of lipopolysaccharide (LPS) in asthma and other pulmonary conditions. *J. Endotoxin Res.* 9:293–300.
- Monroe, S., and R. Polk. 2000. Antimicrobial use and bacterial resistance. *Curr. Opin. Microbiol.* 3:496–501.
- Newby, D.T., I.L. Pepper, and R.M. Maier. 2000. Microbial transport. p. 147–175. *In* R.M. Maier et al. (ed.) *Environmental microbiology*. Academic Press, San Diego, CA.
- NRC. 2002. Biosolids applied to land: Advancing standards and practices. p. 1–12. National Academy Press, Washington, DC.
- Olenchock, S.A. 2001. Airborne endotoxin. p. 814–826. *In* C.J. Hurst et al. (ed.) *Manual of environmental microbiology*. 2nd ed. ASM Press, Washington, DC.
- Paez-Rubio, T., A. Ramarui, J. Sommer, H. Xin, J. Anderson, and J. Peccia. 2007. Emission rates and characterization of aerosols produced during the spreading of dewatered class B biosolids. *Environ. Sci. Technol.* 41:3537–3544.
- Paez-Rubio, T., H. Xin, J. Anderson, and J. Peccia. 2006. Particulate matter composition and emission rates from the disk incorporation of Class B biosolids into soil. *Atmos. Environ.* 40:7034–7045.
- Pancorbo, O.C., P.R. Scheuerman, S.R. Farrah, and G. Bitton. 1981. Effect of sludge type on poliovirus association with and recovery from sludge solids. *Can. J. Microbiol.* 27:279–287.
- Pasquill, F. 1961. The estimation of the dispersion of wind borne material. *Meteorol. Mag.* 90:33–49.
- Pepper, I.L., J.P. Brooks, and C.P. Gerba. 2006. Pathogens in biosolids. p. 1–41. *In* D.L. Sparks (ed.) *Advances in agronomy*. Elsevier Science/Academic Press, San Diego, CA.
- Pepper, I.L., K.L. Josephson, R.I. Bailey, M.D. Burr, and C.P. Gerba. 1993. Survival of indicator organisms in Sonoran Desert soil amended with sewage sludge. *J. Environ. Sci. Health A* 28:1287–1302.
- Pieper, A.P., J.N. Ryan, R.W. Harvey, G.I. Amy, T.H. Illangasekare, and D.W. Metge. 1997. Transport and recovery of bacteriophage PRD1 in a sand and gravel aquifer: Effect of sewage-derived organic matter. *Environ. Sci. Technol.* 31:1163–1170.
- Pillai, S.D., K.W. Widmer, S.E. Dowd, and S.C. Ricke. 1996. Occurrence of airborne bacteria and pathogen indicators during land application of sewage sludge. *Appl. Environ. Microbiol.* 62:296–299.
- Quanrud, D., J. Chorover, and E. Saez. 2007. Polybrominated diphenyl ethers in biosolids: Assessment of relative risk after land application. University of Arizona National Science Foundation Water Quality Center Report, Spring 2007.
- Rensing, C., D.T. Newby, and I.L. Pepper. 2002. The role of selective pressure and selfish DNA in horizontal gene transfer and soil microbial community adaptation. *Soil Biol. Biochem.* 34:285–296.
- Roberts, P., J.P. Roberts, and D.L. Jones. 2006. Behavior of the endocrine disrupting chemical nonylphenol in soil: Assessing the risk associated with spreading contaminated waste to land. *Soil Biol. Biochem.* 38:1812–1822.
- Rusin, P., S. Maxwell, J. Brooks, C. Gerba, and I. Pepper. 2003. Evidence for the absence of *Staphylococcus aureus* in land applied biosolids. *Environ. Sci. Technol.* 37:4027–4030.
- Rylander, R., M. Lundholm, and C.S. Clark. 1983. Exposure to aerosol of microorganisms and toxin during handling of sewage sludge. p. 69–78. *In* P.M. Wallis and D.L. Lehmann (ed.) *Biological health risk of sludge disposal to land in cold climates*. Univ. of Calgary Press, Calgary, Alberta.
- Salyers, A.A., A. Gupta, and Y. Wang. 2004. Human intestinal bacteria as reservoirs for antibiotic resistance genes. *Trends Microbiol.* 12:412–416.
- Smid, T., D. Heederik, R. Houbba, and P.H. Quanjer. 1992. Dust and endotoxin related respiratory effects in the animal feed industry. *Am. Rev. Respir. Dis.* 146:1474–1479.
- Sorber, C.A., B.E. Moore, D.E. Johnson, H.J. Harding, and R.E. Thomas. 1984. Microbiological aerosols from the application of liquid sludge to land. *J. Water Pollut. Control Fed.* 56:830–836.
- Stagg, C.H., C. Wallis, C.H. Ward, and C.P. Gerba. 1978. Chlorination of solids-associated coliphages. *Prog. Water Technol.* 10:381–387.
- Stamatiadis, S., J.W. Doran, and T. Kettler. 1999. Field and laboratory evaluation of soil quality changes resulting from injection of liquid sewage sludge. *Appl. Soil Ecol.* 12:263–272.
- Straub, T.M., I.L. Pepper, and C.P. Gerba. 1993. Virus survival in sewage sludge amended desert soil. *Water Sci. Technol.* 27:421–424.
- Tanner, B.D., J.P. Brooks, C.N. Haas, C.P. Gerba, and I.L. Pepper. 2005. Bioaerosol emission rate and plume characteristics during land-application of liquid class B biosolids. *Environ. Sci. Technol.* 39:1584–1590.
- Thomason, B.M., J.W. Biddle, and W.B. Cherry. 1975. Detection of *Salmonellae* in the environment. *Appl. Microbiol.* 30:764–767.
- Tian, G., T.C. Granato, R.I. Pietz, C.R. Carlson, and Z. Abedin. 2006. Effect of long-term application of biosolids for land reclamation on surface water chemistry. *J. Environ. Qual.* 35:101–113.
- Tsadilas, C.D., I.K. Mitsios, and E. Golia. 2005. Influence of biosolids application on some soil physical properties. *Commun. Soil Sci. Plant Anal.* 36:709–716.
- USEPA. 1994. A plain English guide to the EPA Part 503 Biosolids Rule. EPA 832/R-93/003 USEPA, Washington, DC.
- Wellings, F.M., A.L. Lewis, and C.W. Mountain. 1976. Demonstration of solids-associated virus in wastewater and sludge. *Appl. Environ. Microbiol.* 31:354–358.
- Zaleski, K.J., K.L. Josephson, C.P. Gerba, and I.L. Pepper. 2005a. Survival, growth, and regrowth of enteric indicator and pathogenic bacteria in biosolids, compost, soil, and land applied biosolids. *J. Resid. Sci. Technol.* 2:49–63.
- Zaleski, K.J., K.L. Josephson, C.P. Gerba, and I.L. Pepper. 2005b. Potential regrowth and recolonization of *Salmonella* and indicators in biosolids and biosolid amended soil. *Appl. Environ. Microbiol.* 71:3701–3708.
- Zerzghi, H. 2008. Sustainability of long-term land-application of Class B biosolids: Influence on soil microbial and chemical properties. Ph.D. diss. Univ. of Arizona, Tucson, AZ.